For Research Use Only

anti-Rabbit IgG/anti-Mouse IgG Magnetic VHH Agarose for Immunoprecipitation

Catalog Number: mrIGma

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Basic Information

Catalog Number:
mrIGma

Applications:
IP, Co-IP

Conjugate:
Magnetic Agarose beads; ~40 um (cross-linked 6% magnetic agarose beads)

Host:
Alpaca

Type:
Nanobody

Class:
Recombinant - Animal free production

Description

anti-Rabbit / anti-Mouse IgG IP Beads is an affinity resin for IP of all subtypes of Rabbit and Mouse IgG. It consists of rabbit and mouse IgG specific VHHs (Nanobodies) coupled to Magnetic Agarose beads.

Binding capacity

/

Elution buffer

SDS Sample Buffer

Wash buffer compatibility

/

Affinity (K_D)

/

Storage

Storage:
+4°C / do not freeze!

Storage Buffer:
20% Ethanol

For technical support and original validation data for this product please contact:

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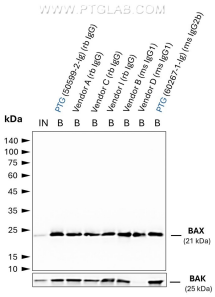
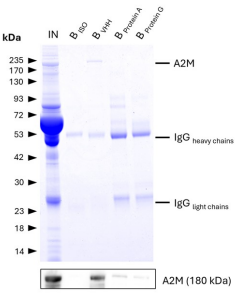
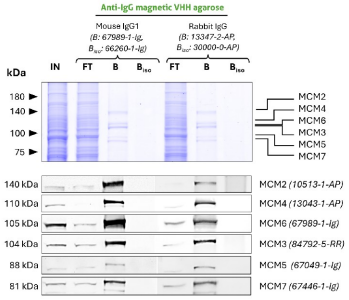
Selected Validation Data



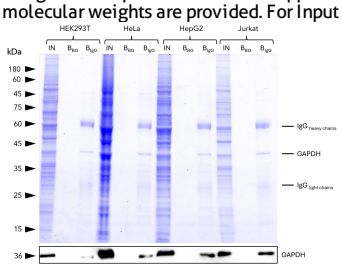
Co-IP of BAX and BAK using anti-BAX IgGs from different vendors by anti-rabbit IgG/ anti-mouse IgG VHH Magnetic Agarose (mrIGma). Tested IgGs include such of rabbit and mouse origin with IgG1 and IgG2b subtypes. 5 µg of respective IgG was spiked into HEK293T cell lysate derived from 0.5x10^7 cells. 1% of input (IN) and 25% of bound (B) fraction was loaded onto an SDS-PAGE gel. For Western Blot analysis BAX was detected using a polyclonal :2000) labeled us P- conjugate(K co- precipitate polyclonal rabbit IgG ile all IgGs allow was absent in the case endor D, possibly d terface being mas



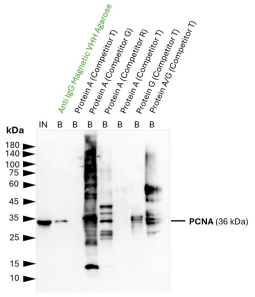
IP of Alpha-2-Macroglobulin (A2M) from human serum using anti-rabbit IgG / anti-mouse IgG magnetic VHH agarose (mrIGma) and comparison to protein A and G beads. IP was performed using 5 µg of rabbit anti-A2M IgG (13545-1-AP) in 1:10 diluted human serum. 0.5% and 6.25% of input (IN) and bound (B) fractions were analyzed by SDS-PAGE, respectively. 5 µg of rabbit isotype control antibody (BISO, 98136-1-RR) was used as negative control. The Coomassie-stained gel shows precipitation of A2M by anti-rabbit IgG / anti-mouse IgG magnetic VHH agarose (BVHH) and reveals its absence for protein A (BPtA) and G (BPtG) beads. This is likely due to human IgGs competing with the spiked IP-antibody. Results were verified using Western blot analysis, which detected A2M at approximately 180 kDa using an anti-A2M primary IgG (13545-1-AP, 1:5000).



Co-IP of MCM complex via pulldown of MCM6 using 5 µg of anti-MCM6 antibodies and anti-rabbit IgG / anti-mouse IgG magnetic VHH agarose (mrIGma). All subunits of the 600 kDa heterohexameric complex are successfully precipitated using both a mouse IgG1 (67989-1-Ig) and a rabbit (13347-2-AP) MCM6 antibody, as shown by western blot analysis using subunit specific antibodies. Apparent molecular weights are provided. For Input (IN) and



IP of GAPDH from common human cell lines using anti-rabbit IgG/ anti-mouse IgG Magnetic VHH Agarose (mrIGma). IP was performed using 5 µg of rabbit anti-GAPDH IgG (10494-1-AP) with 1x107 cells used per IP reaction. 1% and 25% of input (IN) and bound (B) fractions were analyzed by SDS-PAGE, respectively. Bound fractions without antibody spiked (BBG) are shown as background control. The Coomassie-stained gel shows precipitation of GAPDH by anti-rabbit IgG / anti-mouse IgG magnetic VHH agarose, what was verified using Western blot analysis using the anti-GAPDH mouse monoclonal antibody (60004-1-Ig).



IP of PCNA using 5 µg of anti-PCNA antibody (IgG1, 60097-1-Ig) by anti-rabbit IgG/ anti-mouse IgG Magnetic VHH agarose (mrIGma). For Western blot analysis the PCNA polyclonal antibody (10205-2-AP, 1:2000) was labeled using a conformation-specific HRP-conjugated secondary to avoid staining of heavy and light chain of the IP-antibody. In contrast to many competitor protein A and G beads, clean pulldown of PCNA by Proteintech anti-rabbit IgG/ anti-mouse IgG magnetic VHH agarose facilitates unambiguous identification of the target protein. Other beads show leaching of protein A or protein G fragments into the final IP fractions, which can lead to binding of the detection antibody and unspecific signals, as reported previously (Grant et al., 2019, Biiol Proced Online, doi: 10.1186/s12575-019-0095-z). For input (IN) and bound (B) fractions 1% and 30% were loaded, respectively.